

Role of stanniocalcin-1 and proenkephalin-A as novel biomarkers in prediction of newly diagnosed type 2 diabetic patients

Baydaa Ahmed Abed¹ · Isam Noori Salman¹ · Ekhlas Abdallah Hassan²  · Noor Ulhuda G. Mohammed³

Received: 5 November 2023 / Accepted: 9 May 2024 / Published online: 20 May 2024

© The Author(s), under exclusive licence to Research Society for Study of Diabetes in India 2024

Abstract

Background Diabetes mellitus is a chronic metabolic disorder that carries substantial implications for health, social well-being, and economic factors. Stanniocalcin-1 STC-1 is a polypeptide hormone that was initially distinguished as a controller of calcium/phosphate homeostasis in the fish. There is a growing evidence that human STC-1, an autocrine/paracrine factor, plays a role in both inflammation and carcinogenesis. The stable fragment proenkephalin-A (PENKA) is a novel biomarker derived from the precursor enkephalin, which is involved in multiple biological pathways.

Objective The objective of this study was to assess the significance of two newly discovered biomarkers, stanniocalcin-1 and proenkephalin-A, in patients recently diagnosed with type 2 diabetes.

Methods The study consisted of 90 participants, who were divided into two groups. The first group comprised 60 patients with recently diagnosed T2DM, while the second group consisted of 30 healthy individuals serving as controls. The age range of the participants spanned from 35 to 65 years.

Result The study found a significant increase in body mass index (BMI), fasting blood glucose (FBG), HbA1c, C-peptide, HOMA-IR, total cholesterol (TC), triglyceride (TG), LDL, VLDL, stanniocalcin-1, and proenkephalin-A in patients with newly DM when compared with those in the control group. Also, there was a significant increase in HDL levels in the control group compared to the newly diagnosed diabetic group.

Conclusions In conclusion, the results of our study indicate that stanniocalcin-1 and proenkephalin-A could potentially be used as indicators of glucose homeostasis. Consequently, these biomarkers may offer an alternative to conventional markers in predicting risk factors for diabetes mellitus.

Keywords Diabetes mellitus · Stanniocalcin-1 · Proenkephalin-A · Lipid profile · HOMA-IR

Introduction

Diabetes mellitus (DM) is a chronic endocrine disease that is characterized by high blood sugar levels. This condition occurs due to either insufficient production of insulin by the pancreas or reduced sensitivity of the body to insulin, or both [1]. Insufficient activity of insulin on target tissue, due to insulin insensitivity or absence, leads to disturbances in carbohydrate, protein, and fat metabolism. Type 2 diabetes affects individuals of various age groups and has significant implications for morbidity, mortality, and healthcare expenses for patients, their families, and countries. A study conducted in 2013 revealed that it impacted 382 million people (7.7% of the population), with projections estimating a continuous increase to 483 million (8.3%) by the year 2030. In developed nations, a majority of individuals diagnosed with T2DM are over the age of 65, while only 8% are under

✉ Ekhlas Abdallah Hassan
ekhlasbiochemistry@gmail.com

Baydaa Ahmed Abed
baydaaahmed@yahoo.com

Isam Noori Salman
esamnoori61@gmail.com

Noor Ulhuda G. Mohammed
noorulhuda.g@csu.uobaghdad.edu.iq

¹ National Diabetes Center, Mustansiriyah University, Baghdad, Iraq

² Department of Chemistry, College of Science, Diyala University, Baquba, Diyala, Iraq

³ Department of Chemistry, College of Science for Women, University of Baghdad, Al-Jadriya, Baghdad, Iraq

the age of 44 [2]. Insulin resistance characterization of type 2 diabetic is associated with a progressive decay of β -cell function in pancreas. T2DM is additionally connected to an expanded danger of cardiovascular comorbidities and cardiovascular disease [3]. Stanniocalcin-1 STC-1 is an auto-crine/paracrine factor in mammalian that regulates calcium/phosphate homeostasis and was first segregated in fish. The STC hormone family, which consists of STC-1 and STC-2, is commonly expressed in mammalian tissues. However, limited knowledge exists regarding the impact of STC on lipid metabolism [4, 5]. Mammalian STC-1 is communicated in different tissues, including hormone responsive organs and the endocrine glands. Among all tissues, the ovaries contain the most levels of stanniocalcin-1 expression, with raised expression observed during lactation and pregnancy [4]. Furthermore, STC expression has been confirmed in the tissue of pancreas; in fact, STC-1 has described to be expressed together with insulin in the β cells of the pancreas islets [6]. STC is found in various bodily tissues of animals and acts as a paracrine/autocrine factor. Recent research has indicated that the expression of STC-1 is associated with several pathological processes and diseases, including tumor development, inflammation, and metastasis [5]. Proenkephalin-A PENKA is a peptide (approximately 4.5 kDa) [7, 8]. The human proenkephalin is composed of 267 amino acids. It is found in various locations within the nervous system, including the adrenal medulla, bone-derived cells, and different cells of the immune system [9, 10]. Pancreatic islets cells additionally generate proenkephalins, which block the absorption of glucose and hence decrease the insulin response. Moreover, several additional physiological impacts have been documented, such as its impact on blood pressure and heart rate [11]. Plasma can be used to quantify PENKA, a reliable and stable substitute for the enkephalins. The precursor molecule proenkephalin is synthesized in various cells throughout the human body, including both neuronal and non-neuronal cells [12]. The precursor molecule, proenkephalin, is synthesized in various cells within the human body, both neuronal and non-neuronal. The study is aim to assess the significance of two newly discovered biomarkers, stanniocalcin-1 and proenkephalin-A, in patients recently diagnosed with type 2 diabetes.

Material and methods

Study design

A case control study was conducted with a total of 90 participants, based on ADA criteria for diagnosis of DM 2022 [13]. The first group consisted of 60 individuals who had recently been diagnosed with T2DM (newly DM), while the second group included 30 healthy subjects. The age range

of the participants was between 35 and 65 years, and the healthy subjects were matched in age to the control group for comparison. The exclusion criteria included the following: patients with diabetic group have no other disease like hypertension, ischemic heart disease, and thyroid dysfunction. The study, which took place at the national diabetes center of Mustansiriyah University from July to September 2023.

Ethical approval

The research was conducted in line with the Helsinki Declaration. Obtained informed consent from all participants prior to their involvement. Furthermore, the study received ethical approval from the ethics committee of the national diabetes center.

Collection of a blood samples

The samples were collected from individuals who had fasted for 8–12 h between the hours of 8:30 and 11:30 a.m. The blood was divided into two parts. One part, consisting of 1 mL, was transferred into an EDTA tube for the measurement of HbA1C. The other part, consisting of 9 mL, was transferred to a gel tube. Two milliliters of serum from this part were used for the measurement of FBG and lipid profile. The remaining portion was transferred to a tube and stored at -20°C for further investigation, specifically C-peptide, stanniocalcin-1, and proenkephalin-A.

Anthropometries measurement

We measured age, weight, and height of the participants. The BMI was calculated by dividing the weight (in kilograms) by the square of the height (in meters).

Measurement of the homeostasis model assessment (HOMA-IR)

Various methods were employed to evaluate insulin resistance (IR) in the research, with the prevailing method being the utilization of homeostasis model assessment (HOMA). HOMA involves the computation of fasting insulin (U/ml) and glucose (mg/dl), as depicted in the subsequent equation [14].

$$\text{HOMA/IR} = [\text{g(mg/dl)} \times \text{fasting insulin } (\mu\text{U/ml})]/405$$

Clinical laboratory analysis of groups

The FBG, lipid profile tests (Biolabo, France) were assessed using an enzymatic technique. The HLC-723GX was based

on the high-performance liquid chromatography HPLC was used to determine HbA1c. Enzyme-linked immune sorbent assay (ELISA) was used to assess C-peptide (Human, USA), stanniocalcin-1, and proenkephalin-A (Al-shkairate-Jordon).

Statistical analysis

The statistical analysis was conducted using SPSS 22.0.0 (2018), Chicago, IL, USA. The significance level was set at $p < 0.05$. Unpaired t -tests were employed to examine differences among multiple groups, assuming a normal distribution and the absence of significant outliers. To assess the ability of various parameters to distinguish active cases from control, receiver operator characteristic curves (ROC) were utilized.

Result

Anthropometrics and biochemical parameters of study groups

The level of (BMI, FBG, HbA1c, C-peptide, and HOMA-IR) showed a highly significant increase ($p < 0.001$) in newly diagnosed DM as compared to control group as illustrated in Table 1.

The result of the study as shown in the Table 2, demonstrated a highly increase significant ($p < 0.001$) in newly diagnosed DM (TC, TG, LDL, and VLDL) as compared to control group. Also, the current study revealed a statistically significant increase ($p < 0.001$) in the mean levels of HDL in the control group. Conversely, the mean levels of HDL were lower in the newly diagnosed DM group.

Serum stanniocalcin-1 and proenkephalin-A of study groups

Serum (stanniocalcin-1 and proenkephalin-A) was significantly higher $p < 0.001$ in newly diagnosed DM

Table 1 Anthropometrics and biochemical parameters of study groups

Variables	Newly DM ($N=60$)	Control ($N=30$)	p -value
Age years	52.4 ± 8.9	50.1 ± 9.4	NS
BMI kg/m ²	32.1 ± 7.1	25.8 ± 4.1	<0.001
FBG (mg/dl)	200.2 ± 50.15	88.6 ± 7.8	<0.001
HbA1c %	8.6 ± 1.2	4.8 ± 0.6	<0.001
C-peptide (ng/ml)	2.3 ± 0.7	1.7 ± 0.6	<0.001
HOMA-IR	3.8 ± 1.8	2.2 ± 0.5	<0.001

All data represent as mean ± SD. BMI body mass index, FBS fasting blood glucose

Table 2 Assessment of lipid panel biochemical parameters of study groups

Variables	Newly DM ($N=60$)	Control ($N=30$)	p -value
TC (mg/dl)	176.9 ± 30.3	140.8 ± 20.5	<0.001
TG (mg/dl)	155.0 ± 30.8	88.6 ± 25.9	<0.001
HDL (mg/dl)	39.7 ± 8.5	47.1 ± 5.8	<0.001
LDL (mg/dl)	109.2 ± 33.5	84.1 ± 24.3	<0.001
VLDL (mg/dl)	28.7 ± 12.6	16.2 ± 3.9	<0.001

All data represent as mean ± SD. TC total cholesterol, TG triglyceride

(1200.2 ± 353.2 and 737.8 ± 254.4) compared to control group (701.6 ± 223.1 and 484.3 ± 283.2).

Receiver operating characteristic curve analysis (ROC)

Stanniocalcin-1 demonstrated significant capability when comparing newly diagnosed DM with the control group, exhibiting high sensitivity and specificity ($\geq 90\%$), as well as high accuracy ($\geq 80\%$). Proenkephalin-A also exhibited a favorable ability to distinguish between newly diagnosed DM and the control group, with high sensitivity and specificity ($\geq 80\%$) and accuracy ($\geq 80\%$), as indicated in Table 3 and Figs. 1 and 2.

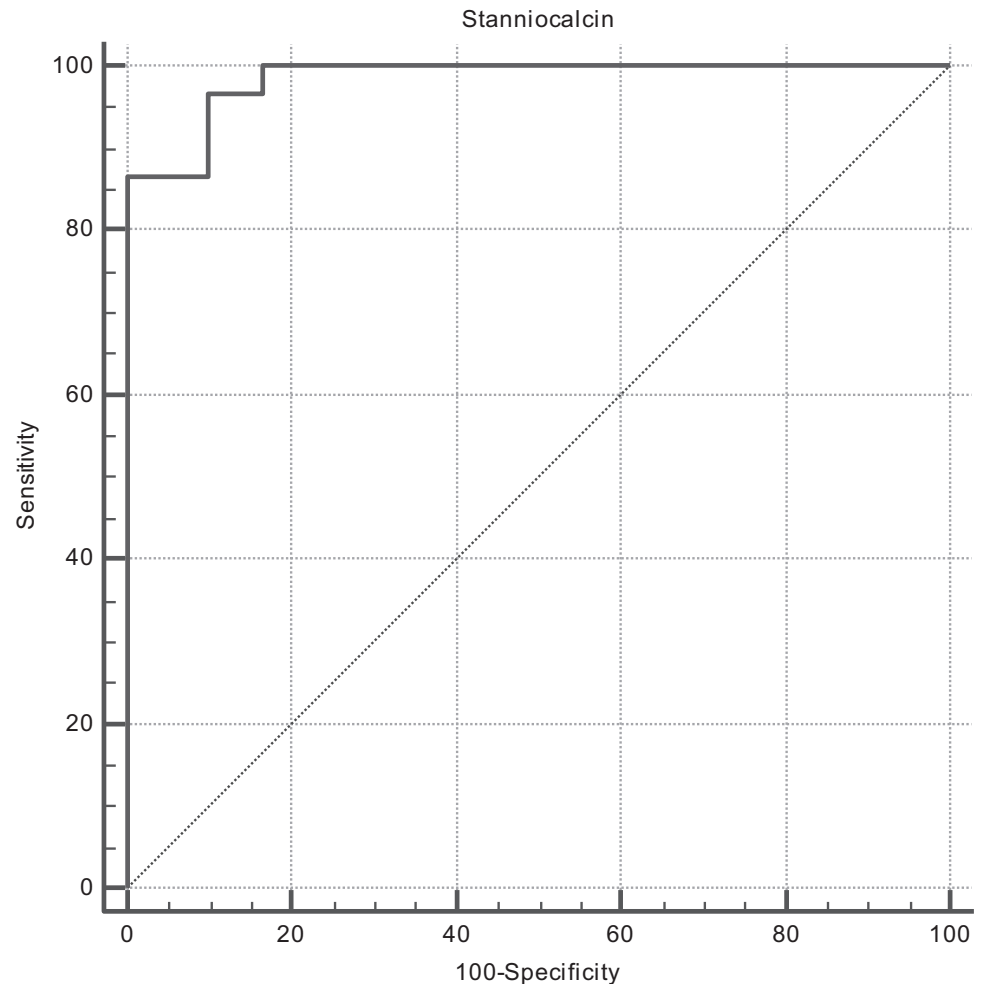
Discussion

Glycated hemoglobin serves as an indicator of blood glucose levels within the preceding 3-month period. In cases where hemolytic anemia medications or kidney dysfunction are present, HbA1c measurements may potentially underestimate the true extent of blood glucose levels [15, 16]. HbA1c testing is significant as it tests chronic glycemia in patients with diabetes. It has been used as an objective criterion for normal glycemic regulation in diabetes patient management [17]. Insulin and connection peptide (C-peptide) are secreted by the pancreatic β cells [18, 19]. C-peptide, especially after glucagon stimulation or a mixed meal, is considered a reliable tool for estimating the residual function of β cells [18, 20]. According to numerous studies, C-peptide is a biologically active peptide. It has been proven to mitigate the inflammation resulting from hyperglycemia and provide protection against diabetic complications in individuals with diabetes mellitus (DM) [20–22]. The findings of this study are consistent with our previous research [22–24], which observed elevated levels of TC, TG, and LDL, as well as reduced levels of HDL cholesterol, among T2DM. By reducing glucose absorption in peripheral tissues and boosting glucose synthesis in the liver, insulin resistance causes high blood glucose levels. Insulin resistance induces heightened

Table 3 AUC and validity of stanniocalcin-1 and proenkephalin-A to differentiate between patients and control groups

Variables	Cut-off value	AUC	SN	SP	AC	PPV	NPV
Stanniocalcin-1	< 897.5	0.974	96.8	90.5	93.7	90.8	97.4
Proenkephalin-A	< 767. <	0.850	83.1	83.2	83.2	83.2	83.2

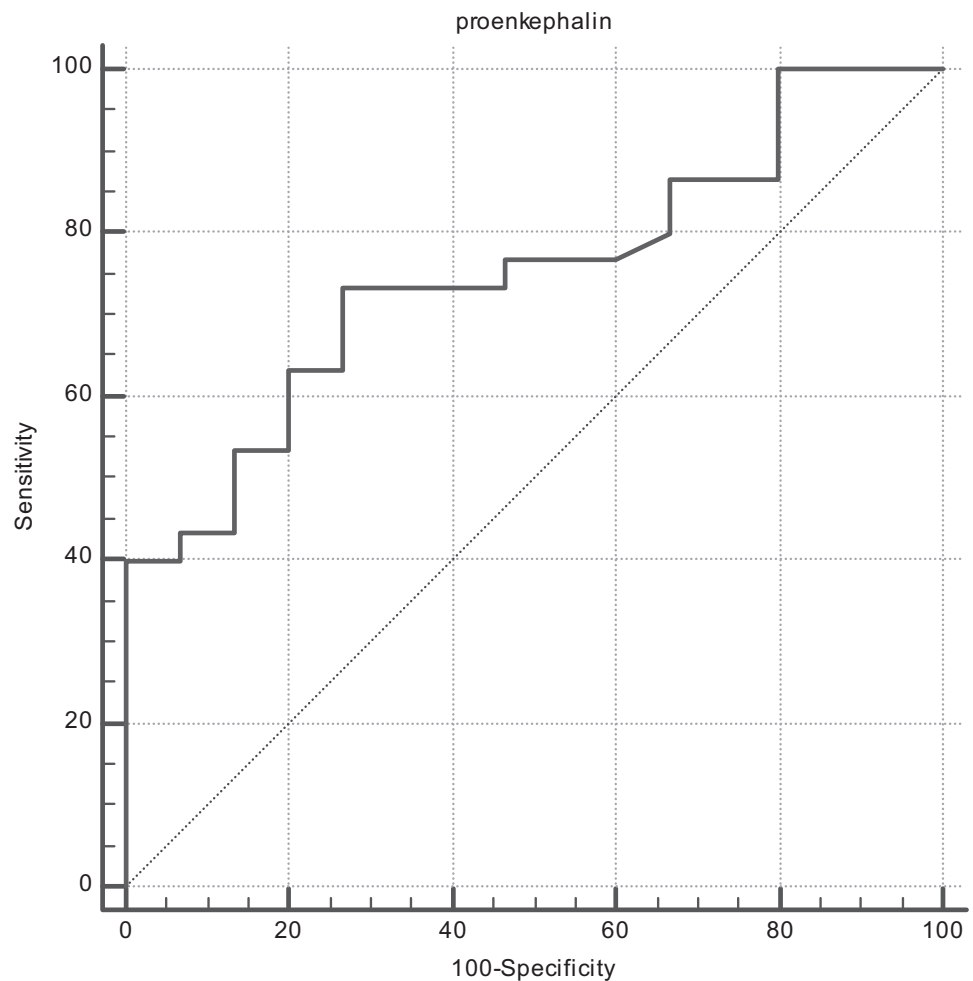
AUC area under the curve, SN sensitivity, SP specificity, AC accuracy, PPV positive predictive value, NPV negative predictive value

Fig. 1 The ROC curve for stanniocalcin-1 newly diagnosed DM with control group

adipocyte lipolysis and the subsequent release of free fatty acids, along with increased hepatic lipid production. Mammalian stanniocalcin-1 STC-1 is a glycoprotein hormone with antioxidant and anti-inflammatory properties that is involved in calcium control. STC-1 has recently been shown to attenuate the development of ROS and the activation of inflammatory pathways by inducing mitochondrial [25–27]. The binding of STC-1 to the cell surface leads to the activation of mitochondria. Based on subcellular fractionation, it is indicated that more than 90% of cellular STC-1 exhibits immunoreactivity in the mitochondrial fraction [23, 28, 29]. The reduction of hypercalcemia tends to be the main feature of STC, close to that of calcitonin. STC and calcitonin both

possess antihypercalcemic properties, although STC exhibits a greater efficacy compared to calcitonin. Furthermore, the expression of human STC-1 is not limited to a specific organ; rather, it is predominantly expressed in various tissues including the thyroid gland, ovary, and prostate [30]. Zaidi, D. et al. [31], showed that STC-1 and insulin have collocated strongly in adult mouse pancreatic β cells. Since the majority of the cellular insulin is restricted to secretory vesicles, much of the immunoreactivity of STC-1 with insulin appears to exist therein. The STC-1 transcript receptor is predominantly located in the β cells of the pancreas, suggesting its involvement in intracrine signaling. However, the specific functions of this receptor in β -cells remain

Fig. 2 The ROC curve for proenkephalin-A in newly diagnosed DM with control groups



speculative [32]. Sarapio, E. et al. [33] showed that human STC-1 could be new signaling elements for the regulation of blood glucose levels in the postprandial period by rising glucose uptake in adipose tissue. In addition, Schein, V. et al. [34] found that STC-1 reduced renal gluconeogenic activity in rats, indicating that these hormones are involved in mammalian homeostasis of blood glucose. The decrease in gluconeogenesis in the kidneys and the rise in glucose absorption by STC-1 hormones in adipose tissue play a crucial role in maintaining glucose balance in mammals, particularly individuals with T2DM. The precursor molecule of the enkephalin family is proenkephalin-A [35, 36]. It is located mostly in the cell matrix and is also present in the nucleus, membrane, and mitochondria of the cell. It also serves as a neurotransmitter, encoding a preproprotein that was proteolytically processed to generate multiple kinds of protein products [37, 38]. The peptides derived from proenkephalin-A function as neuromodulators, neurotransmitters, and neurohormones. They demonstrate opioid activity and have effects on pain responses, stress responses, sleep, and appetite control [39]. The opioid peptide enkephalin is found in brain and endocrine tissues and has effects on nociceptive

and neuroendocrine responses [40]. Enkephalin is produced by cells in the pancreatic islets and functions to inhibit insulin response by blocking glucose absorption. Furthermore, enkephalin has been found to have various physiological effects, including its influence on blood pressure and heart rate [41]. The effects of enkephalin and morphine were blocked by the specific opiate antagonist naloxone hydrochloride (1.2×10^{-6} mol/l). The insulin secretory response of perfused islets to enkephalins and morphine was rapid, corresponding to the first phase of glucose induced insulin release. These observations suggest that there may be opiate receptors in islets, and that opioid peptides could modulate insulin release [11]. The direct effects of an enkephalin analogue on insulin release from isolated islets of Langerhans of the rat have been investigated. It had a dose-dependent effect on insulin secretion [42]. Opioid peptides are localized within the hypothalamus and, thus, are likely involved in modulating homeostatic signals received from the peripheral tissues, pharmacological studies have shown that opioid receptor agonists stimulate food intake, while opioid receptor antagonists suppress food intake. However, the role of each opioid peptide in feeding behavior and, notably,

glucose homeostasis is not fully understood [43]. Marino, R. et al., Shah, KS. et al., Hollinger, A. et al., and Beunders, R. et al. [44–47] found that plasma proenkephalin is elevated in patients with acute or chronic renal failure compared to healthy controls. The proposed pathophysiological pathway for proenkephalin may have prognostic value in predicting mortality and worsening renal function. This pathway could be attributed to the cardio depressive effects of enkephalins, which may lead to decreased kidney perfusion and advanced heart failure. It is difficult to explain properly the real mechanism through which the biomarkers stanniocalcin-1 and proenkephalin-A affect the immune process and glucose homeostasis in diabetic patients and some information may be missed because these markers are novel and there is a lack of references and researchers about them which make it difficult to understand their clear role in pathophysiology. There are certain limitations, it is a cross-sectional study with no follow-up conducted at a single center, and a bit small sample size.

Conclusion

Our results reveal a potential role of proenkephalins and STC-1 in the regulation of glucose homeostasis and in the pathophysiology of diabetes type 2. Stanniocalcin-1 and proenkephalin-A are the most specific and sensitive markers in a newly diagnosed type 2 diabetic. Stanniocalcin-1 and proenkephalin-A are a sign of glucose homeostasis, and hence may serve as a substitute for all other conventional biomarkers in the prediction of risk factors for diabetes mellitus. Our suggestion of measurement of these biomarkers may have utility as novel biomarkers of the newly diagnosed type 2 diabetic.

Author contributions The authors contributed to this work.

Declarations

Ethics approval The research was conducted in line with the Helsinki Declaration. Obtained informed consent from all participants prior to their involvement. Furthermore, the study received ethical approval from the ethics committee of the national diabetes center.

Conflict of interests The authors declare no competing interests.

References

1. Ishii H, Shuichi S, Williams P, Demiya S, Aranishi T, Treuer T. Cross-sectional survey in patients with type 1 and type 2 diabetes to understand mealtime insulin unmet needs in Japan: the MINUTES-J study. *Diabetes Res Clin Pract.* 2020;162:108076.
2. Asiimwe D, Mauti GO, Kiconco R. Prevalence and risk factors associated with type 2 diabetes in elderly patients aged 45–80 years at Kanungu District. *J Diabetes Res.* 2020;2020:1–5.
3. Cassano V, Leo A, Tallarico M, Nesci V, Cimellaro A, Fiorentino TV, Citraro R, Hribal ML, De Sarro G, Perticone F. Metabolic and cognitive effects of ranolazine in type 2 diabetes mellitus: data from an in vivo model. *Nutrients.* 2020;12:382.
4. Chen F, Zhang Z, Pu F. Role of stanniocalcin-1 in breast cancer. *Oncol Lett.* 2019;18:3946–53.
5. Leung CCT, Wong CKC. Effects of STC1 overexpression on tumorigenicity and metabolism of hepatocellular carcinoma. *Oncotarget.* 2018;9:6852.
6. López JJ, Jardín I, Cantonero Chamorro C, Duran ML, Tarancon Rubio MJ, Reyes Panadero M, Jiménez F, Montero R, González MJ, Martínez M. Involvement of stanniocalcins in the deregulation of glycaemia in obese mice and type 2 diabetic patients. *J Cell Mol Med.* 2018;22:684–94.
7. Beunders R, Struck J, Wu AHB, Zarbock A, Di Somma S, Mehta RL, Koyner JL, Nadim MK, Maisel AS, Murray PT. Proenkephalin (PENK) as a novel biomarker for kidney function. *J Appl Lab Med.* 2017;2:400–12.
8. Mohammed NUG, Khaleel FM, Gorial FI. The role of serum chitinase-3-like 1 protein (YKL-40) level and its correlation with proinflammatory cytokine in patients with rheumatoid arthritis. *Baghdad Sci J.* 2022;19:1014.
9. Ernst A, Köhrle J, Bergmann A. Proenkephalin A 119–159, a stable proenkephalin A precursor fragment identified in human circulation. *Peptides.* 2006;27:1835–40.
10. Taha EM, Mohialdeen Taha M, Al-Obaidy SK, Faris Hasan B, Rahim SM. Association between atherogenic index and cholesterol to HDL ratio in COVID-19 patients during the initial phase of infection. *Arch Razi Inst.* 2022;77:1303–10.
11. Green IC, Perrin D, Pedley KC, Leslie RDG, Pyke DA. Effect of enkephalins and morphine on insulin secretion from isolated rat islets. *Diabetologia.* 1980;19:158–61.
12. Schulz C-A, Christensson A, Ericson U, Almgren P, Hindy G, Nilsson PM, Struck J, Bergmann A, Melander O, Orho-Melander M. High level of fasting plasma proenkephalin-A predicts deterioration of kidney function and incidence of CKD. *J Am Soc Nephrol JASN.* 2017;28:291.
13. Draznin B, Aroda VR, Bakris G, Benson G, Brown FM, Freeman R. G. American Diabetes Association Professional Practice Committee, American Diabetes Association Professional Practice Committee. 15. Management of diabetes in pregnancy: standards of medical care in diabetes-2022. *Diabetes Care.* 2022;45:S60–82.
14. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and β -cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia.* 1985;28:412–9.
15. Elghoneimy YF, Nashy MR, Othman SA, Almarri NM, Alruwaili AA, Alotaibi AR, Alshammari FA. Which level of preoperative glycated haemoglobin (HbA1c) affect early morbidity and mortality after cardiac surgery? *Australas Med J.* 2020;13:32–40.
16. Farhan LO, Taha EM, Farhan AM. A Case control study to determine Macrophage migration inhibitor, and N-telopeptides of type I bone collagen Levels in the sera of osteoporosis patients. *Baghdad Sci J.* 2022;19(4):848–54.
17. d’Emden M. Glycated haemoglobin for the diagnosis of diabetes. *Aust Prescr.* 2014;37(3):98–100.
18. Salih JM, Abdulateef DS. Detection of C-peptide in human hair and nail: a comparison between healthy persons and persons with type 1 diabetes. *BMJ Open Diabetes Res Care.* 2020;8:e001297.
19. Kashi GJ, Abed BA, Farhan LO, Noori I, Salman ASD. A Comparative study to determine LDH enzyme levels in serum samples of women with breast cancer and women with breast cancer and type 2 diabetes mellitus. *J Med Chem Sci.* 2023;6:883–90.

20. Araujo DB, Dantas JR, Silva KR, Souto DL, Pereira M de FC, Moreira JP, Luiz RR, Claudio-Da-Silva CS, Gabbay MAL, Dib SA. Allogenic adipose tissue-derived stromal/stem cells and vitamin D supplementation in patients with recent-onset type 1 diabetes mellitus: a 3-month follow-up pilot study. *Front Immunol.* 2020;11:993.
21. Yan S-T, Sun J, Gu Z-Y, Miao X-Y, Ma L-C, Sun B-R, Fu X-M, Liu H-Z, Yang G, Fang F-S. The bidirectional association of C-peptide with cardiovascular risk in nondiabetic adults and patients with newly diagnosed type 2 diabetes mellitus: a retrospective cohort study. *Cardiovasc Diabetol.* 2022;21:201.
22. Abed BA, Al-Aaraji SB, Salman IN. Estimation of Galanin hormone in patients with newly thyroid dysfunction in type 2 diabetes mellitus. *Biochem Cell Arch.* 2021;21:1317–21.
23. Farhan LO, Abed BA, Dawood A. Comparison study between adipsin levels in sera of Iraqi patients with diabetes and neuropathy. *Baghdad Sci J.* 2023;20:726.
24. Abed BA, Hamid GS. Evaluation of lipocalin-2 and vaspin levels in Iraqi women with type 2 diabetes mellitus. *Iraqi J Sci.* 2022;63:4650–8.
25. Khatun M, Arffman RK, Lavogina D, Kangasniemi M, Laru J, Ahtikoski A, Lehtonen S, Paulson M, Hirschberg AL, Salumets A. Women with polycystic ovary syndrome present with altered endometrial expression of stanniocalcin-1. *Biol Reprod.* 2020;102:306–15.
26. Roddy GW, Rinkoski TA, Monson KJ, Chowdhury UR, Fautsch MP. Stanniocalcin-1 (STC-1), a downstream effector molecule in latanoprost signaling, acts independent of the FP receptor for intraocular pressure reduction. *PLoS ONE.* 2020;15:e0232591.
27. Shi M, Yuan Y, Liu J, Chen Y, Li L, Liu S, An X, Luo R, Long D, Chen B. MSCs protect endothelial cells from inflammatory injury partially by secreting STC1. *Int Immunopharmacol.* 2018;61:109–18.
28. Huang L, Belousova T, Chen M, DiMattia G, Liu D, Sheikh-Hamad D. Overexpression of stanniocalcin-1 inhibits reactive oxygen species and renal ischemia/reperfusion injury in mice. *Kidney Int.* 2012;82:867–77.
29. Mohammed NUG, Gorial FI, Khaleel FM, Abed BA, Mutar SA, Farhan LO, Salman IN. Role of human β -defensin-3 in rheumatoid arthritis: an observational single-center study. *Al-Rafidain J Med Sci (ISSN 2789-3219).* 2023;5:S71-75.
30. Ishibashi K, Imai M. Prospect of a stanniocalcin endocrine/paracrine system in mammals. *Am J Physiol Physiol.* 2002;282:F367–75.
31. Zaidi D, Turner JK, Durst MA, Wagner GF. Stanniocalcin-1 colocalizes with insulin in the pancreatic islets. *ISRN Endocrinol.* 2012;2012:834359.
32. Ellard JP, McCudden CR, Tanega C, James KA, Ratkovic S, Staples JF, Wagner GF. The respiratory effects of stanniocalcin-1 (STC-1) on intact mitochondria and cells: STC-1 uncouples oxidative phosphorylation and its actions are modulated by nucleotide triphosphates. *Mol Cell Endocrinol.* 2007;264:90–101.
33. Sarapio E, De Souza SK, Model JFA, Trapp M, Da Silva RSM. Stanniocalcin-1 and -2 effects on glucose and lipid metabolism in white adipose tissue from fed and fasted rats. *Can J Physiol Pharmacol.* 2019;97:916–23.
34. Schein V, Kucharski LC, Guerreiro PMG, Martins TL, Morgado I, Power DM, Canario AVM, da Silva RSM. Stanniocalcin 1 effects on the renal gluconeogenesis pathway in rat and fish. *Mol Cell Endocrinol.* 2015;414:1–8.
35. Khorashadi M, Beunders R, Pickkers P, Legrand M. Proenkephalin: a new biomarker for glomerular filtration rate and acute kidney injury. *Nephron.* 2020;144:655–61.
36. Holm H, Nägga K, Nilsson ED, Ricci F, Melander O, Hansson O, Bachus E, Fedorowski A, Magnusson M. High circulating levels of midregional proenkephalin A predict vascular dementia: a population-based prospective study. *Sci Rep.* 2020;10:8027.
37. Tang D, Lin T, Wang Y, Cao H. High expression of proenkephalin is associated with favorable outcomes in patients with gastrointestinal stromal tumors. *Cancer Manag Res.* 2019;11:6681–90.
38. Mohammed NUG, Khaleel FM, Gorial FI. Cystatin D as a new diagnostic marker in rheumatoid arthritis. *Gene Reports.* 2021;23:101027.
39. Le Merrer J, Becker JA, Befort K, Kieffer BL. Reward processing by the opioid system in the brain. *Physiol Rev.* 2009;89:1379–412.
40. Vuong C, Van Uum SHM, O'Dell LE, Lutfy K, Friedman TC. The effects of opioids and opioid analogs on animal and human endocrine systems. *Endocr Rev.* 2010;31:98–132.
41. van Hateren KJJ, Landman GWD, Arnold JFH, Joosten H, Groenier KH, Navis GJ, Sparwasser A, Bakker SJL, Bilo HJG, Kleefstra N. Serum proenkephalin A levels and mortality after long-term follow-up in patients with type 2 diabetes mellitus (ZODIAC-32). *PLoS ONE.* 2015;10:e0133065.
42. Khawaja XZ, Green IC, Thorpe JR, Bailey CJ. Increased sensitivity to insulin-releasing and glucoregulatory effects of dynorphin A 1–13 and U 50488h in ob/ob versus lean mice. *Diabetes.* 1990;39:1289–97.
43. Gupta A, Gullapalli S, Pan H, Ramos-Ortolaza DL, Hayward MD, Low MJ, Pintar JE, Devi LA, Gomes I. Regulation of opioid receptors by their endogenous opioid peptides. *Cell Mol Neurobiol.* 2021;41:1103–18.
44. Marino R, Struck J, Hartmann O, Maisel AS, Rehfeldt M, Magrini L, Melander O, Bergmann A, Di Somma S. Diagnostic and short-term prognostic utility of plasma pro-enkephalin (pro-ENK) for acute kidney injury in patients admitted with sepsis in the emergency department. *J Nephrol.* 2015;28:717–24.
45. Shah KS, Taub P, Patel M, Rehfeldt M, Struck J, Clopton P, Mehta RL, Maisel AS. Proenkephalin predicts acute kidney injury in cardiac surgery patients. *Clin Nephrol.* 2015;83:29–35.
46. Hollinger A, Wittebole X, François B, Pickkers P, Antonelli M, Gayat E, Chousterman BG, Lascarrou J-B, Dugernier T, Di Somma S. Proenkephalin A 119–159 (Penkid) is an early biomarker of septic acute kidney injury: the kidney in sepsis and septic shock (Kid-SSS) study. *Kidney Int Rep.* 2018;3:1424–33.
47. Beunders R, van Groenendaal R, Leijte GP, Kox M, Pickkers P. Proenkephalin compared to conventional methods to assess kidney function in critically ill sepsis patients. *Shock.* 2020;54:308.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.